Amendments to the Specification

Please amend the paragraph beginning on page 24, line 26, as follows:

Method: Twelve plants of 96SN-0510 were planted in the growth room under controlled environment as donor plants. These plants were sprayed with the herbicide, PURSUIT™ (imazethapyr), at 1x level. Immature buds were harvested from each donor plant and were crushed in a blender to produce a slurry [as described in Swanson, E.B. et al., "Efficient isolation of microspores and the production of microspore-derived embyros in *Brassica napus*", L. Plant Cell Reports, 6: 94-97, (1987); and Swanson, E.B., Microspore culture in *Brassica*, pp. 159-169 In Methods in Molecular Biology, vol. 6, Plant Cell and Tissue Culture, Humana Press, (1990)]. The slurry was then filtered through two layers of Nitex filters (48 μm pores) and collected in centrifuge tubes. The suspensions were centrifuged, decanted and washed three times for a total of 4 spins. Microspores were counted using a haemocytometer and plated in NLN medium (Lichter, R. "Induction of haploid plants from isolated pollen of *Brassica napus*", Z. Pflanzenphysiol. Bd. 105: 427-434, ~~(1982)~~(1982)), containing 40μg/l PURSUIT™, at a density of 60,000 microspores per ml. Ten ml of this suspension was poured into 100x25mm petri plates, wrapped with parafilm, and placed in a Percival incubation chamber at 32.5°C in darkness for 15 days. During this period, the microspores carrying the genes responsible for resistance to Imidazolinone herbicides were expected to survive and produce embryos. After 15 days, petri plates with cotyledonary embryos were put on a rotary shaker for 6 to 13 days before being transferred to solid 0.8% agar medium with 0.1% Gibberillic acid

(GA) in petri plates. Transferred embryos were incubated in the dark at 4-8^{0}C for 7 to 10 days and removed to a Percival incubation chamber in light at 20 to 25^{0}C for 3 to 5 weeks. Selected embryos that regenerated were placed in soil in 72 cell flats or put back onto 0.8% agar with 0.1% GA for a further 3 to 5 weeks before they were transplanted to soil. Before flowering, plants were treated with 0.33% colchicine for 1.5 to 2.5 hours. Plant roots were washed free of soil prior to incubation in the colchicine solution. After treatment they were planted in 10 cm plastic pots. Upon flowering, plants with fertile (diploid) racemes were covered with perforated, clear plastic bags to produce selfed seeds. After flowering bags were removed and plants were dried down, seeds was harvested, cleaned, and catalogued with a DHS number. Lines with 100 seeds or more were prepared for nursery evaluation.